MycoKeys 85: 73–85 (2021)
doi: 10.3897/mycokeys.85.76040
https://mycokeys.pensoft.net

RESEARCH ARTICLE

A peer-reviewed open-access journal
MycoKeys
Launched to accelerate biodiversity research

# New species and records of *Chapsa* (Graphidaceae) in China

Ming-Zhu Dou[1*], Min Li[1*], Ze-Feng Jia[1]

1 *College of Life Sciences, Liaocheng University, Liaocheng 252059, China*

Corresponding author: Ze-Feng Jia (zfia2008@163.com)

Academic editor: Thorsten Lumbsch | Received 5 October 2021 | Accepted 27 November 2021 | Published 10 December 2021

**Citation:** Dou M-Z, Li M, Jia Z-F (2021) New species and records of *Chapsa* (Graphidaceae) in China MycoKeys 85: 73–85. https://doi.org/10.3897/mycokeys.85.76040

## Abstract

We studied the genus *Chapsa* in China based on morphological characteristics, chemical traits and molecular phylogenetic analysis. One species new to science (*C. murioelongata* M.Z. Dou & M. Li) and two records new to China were found (*C. wolseleyana* Weerakoon, Lumbsch & Lücking and *C. niveocarpa* Mangold). *Chapsa murioelongata* **sp. nov.** is characterised by its lobed thalline margin, orange discs with white pruina, clear hymenium, and submuriform and long ascospores. *Chapsa wolseleyana* was recombined into *Astrochapsa* based on phenotypic traits. Sequences of this species are for the first time reported here and phylogenetic analyses of three loci (mtSSU, ITS and nuLSU) supported the position of this species within *Chapsa*. A key for the *Chapsa* species known in China is provided.

## Keywords

Ascomycota, lichenized fungi, phylogeny, taxonomy

## Introduction

The lichen genus *Chapsa* (Graphidaceae) was first established by Massalongo (1860) with *C. indica* as the type species. This genus was ignored for a long time until 2006, when Frisch re-established *Chapsa*, based on the *Chroodiscus*-type apothecia, presence of periphysoids and *Chapsa*-type paraphyses. Frisch (2006) also provided a detailed description and delimitation of the genus *Chapsa*, which was widely recognised by

* These two authors contributed equally to this work.

Copyright *Ming-Zhu Dou et al.* This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

subsequent researchers (Mangold 2008; Frisch and Kalb 2009; Rivas Plata et al. 2011; Sipman et al. 2012; Xu et al. 2016). The genus *Chapsa* was considered to be monophyletic in the beginning (Frisch 2006) but with further research, it was suspected to be polyphyletic (Mangold 2008; Papong et al. 2010). Subsequently, seven genera, *Astrochapsa* Parnmen, Lücking & Lumbsch, *Crutarndina* Parnmen, Lücking & Lumbsch, *Gintarasia* Kraichak, Lücking & Lumbsch, *Pseudochapsa* Parnmen, Lücking & Lumbsch, *Pseudotopeliopsis* Parnmen, Lücking & Lumbsch, *Myriochapsa* M. Cáceres, Lücking & Lumbsch and *Nitidochapsa* Parnmen, Lücking & Lumbsch were separated from *Chapsa*, based on a combination of molecular evidence, phenotypic and chemical characteristics (Parnmen et al. 2012, 2013; Kraichak et al. 2013).

Although China is rich in lichenised fungal species (Wei 2020), there are few studies and reports on the genus *Chapsa*. More than 60 species of *Chapsa* have been reported in the world, of which only three, *C. indica* A. Massal, *C. mirabilis* A. (Zahlbr.) Lücking and *C. leprocarp*a (Nyl.) Frisch, have so far been found in China (Rivas Plata et al. 2010; Xu et al. 2016; Jia and Lücking 2017; Kalb and Kalb 2017; Wijayawardene et al. 2017; de Lima et al. 2019).

During the study of *Chapsa* A. Massal. in southern China, one species, *C. murioelongata* was found new to science, and two species, *C. niveocarpa* Mangold and *C. wolseleyana* Weerakoon, Lumbsch & Lücking were found new to China. In our study, 26 sequences were newly generated from freshly collected specimens.

## Materials and methods

### Morphological and chemical analyses

The specimens were collected from southern China and deposited in the Fungarium, College of Life Sciences, Liaocheng University, China (LCUF). Morphological and anatomical characters of thalli and apothecia were examined and photographed under an Olympus SZX16 dissecting microscope and an Olympus BX53 compound microscope. The lichen secondary metabolites were detected and identified by thin-layer chromatography using solvent C (Orange et al. 2010; Jia and Wei 2016).

### DNA extraction, PCR sequencing and phylogenetic analysis

Genomic DNA was extracted from ascomata using the Hi-DNA-secure Plant Kit (Tiangen, Beijing, China) according to the manufacturer's protocol. The nuLSU, ITS and mtSSU regions were amplified using the primer pair AL2R/LR6 (Mangold et al. 2008, Vilgalys and Hester 1990), ITS1F/ITS4 (Gardes and Bruns 1993, White et al. 1990) and mrSSU1/mrSSU3R (Zoller et al. 1999), respectively. The PCR amplification progress followed Dou et al. (2018) and the PCR products were sequenced by Biosune Inc. (Shanghai). The newly generated sequences were submitted to GenBank (Table1).

**Table 1.** Information for the sequences used in this study. Newly generated sequences are shown in bold.

| Species | Specimen No. | Locality | ITS | nuLSU | mtSSU |
|---|---|---|---|---|---|
| *Pseudochapsa phlyctidioides* | Lumbsch 20500d | Fiji | – | JX465301 | JX421005 |
| *Pseudochapsa dilatata* | Luecking 32101 | Venezuela | – | JX421446 | JX420981 |
| *Pseudochapsa esslingeri* | Caceres s.n. | Brazil | – | – | JX420983 |
| *Pseudochapsa esslingeri* | Caceres 6006a | Brazil | – | – | JX420984 |
| *Pseudochapsa esslingeri* | Rivas Plata 107C (F) | Peru | – | – | JX420985 |
| *Pseudochapsa esslingeri* | Rivas Plata 809a (F) | Peru | – | – | JX420986 |
| *Chapsa alborosella* | Luecking 31238a | Brazil | – | JX421439 | JX420972 |
| *Chapsa alborosella* | Luecking 25587 | Guatemala | – | JX421440 | JX420973 |
| *Chapsa soredicarpa* | Luecking 31200 | Brazil | – | JX421462 | JX421011 |
| *Chapsa soredicarpa* | Luecking 31240 | Brazil | – | JX421463 | JX421012 |
| *Chapsa sublilacina* | Luecking RLD056 | Mexico | – | HQ639624 | HQ639600 |
| *Chapsa thallotrema* | Lucking 32019 | Venezuela | – | JX465319 | JX421013 |
| *Chapsa indica* | Parnmen018486(RAMK) | Thailand | – | JX465295 | JX465280 |
| *Chapsa leprocarpa* | GZ19531 | China, Guizhou | **MW009079** | **MW007981** | **MW010276** |
| *Chapsa leprocarpa* | GZ19537 | China, Guizhou | **MW009077** | **MW007984** | **MW010278** |
| *Chapsa leprocarpa* | GZ19536 | China, Guizhou | **MW009080** | **MW007982** | **MW010274** |
| *Chapsa niveocarpa* | HN19508 | China, Hainan | **MW009076** | **MW010272** | – |
| *Chapsa niveocarpa* | Lumbsch_19125k2(F) & Mangold (F) | Australia, Queensland | – | – | EU675274 |
| *Chapsa niveocarpa* | Lumbsch 19151p & Mangold (F) | Australia, Queensland | – | FJ708487 | EU075567 |
| *Chapsa patens* | FJ19131 | China, Fujian | **MT995055** | **MW007979** | **MW010275** |
| *Chapsa patens* | FJ19049 | China, Fujian | **MW007918** | **MW007980** | – |
| *Chapsa wolseleyana* | FJ19158 | China, Fujian | **MW009078** | **MW010273** | **MW010277** |
| *Chapsa wolseleyana* | FJ19148 | China, Fujian | **MW009106** | **MW010270** | **MW010279** |
| *Chapsa murioelongata* | HN19222 | China, Hainan | **MW009102** | **MW010271** | – |
| *Chapsa murioelongata* | HN19682 | China, Hainan | **MW009103** | **MW010269** | – |
| *Chapsa pulchra* | CHAPUL19129t | Australia | – | KC020261 | KC020255 |
| *Astrochapsa meridensis* | Luecking 17770 (F) | Costa Rica | – | EU075655 | EU075610 |
| *Astrochapsa mastersonii* | Lumbsch 20500f | Fiji | – | – | JX420996 |
| *Astrochapsa zahlbruckneri* | Papong 6516 | Thailand | – | JX421467 | – |
| *Astrochapsa astroidea* | Lumbsch 19166n & Mangold(F) | Australia, Queensland | – | EU075614 | EU075566 |
| *Astrochapsa astroidea* | Lumbsch 19750a | Thailand | – | JX421441 | JX420974 |
| *Astrochapsa astroidea* | Papong 6004 | Thailand | – | JX421442 | JX420975 |
| *Astrochapsa astroidea* | Luecking 24006 | Thailand | – | JX421443 | JX420977 |
| *Astrochapsa astroidea* | Luecking 24008 | Thailand | – | JX421444 | JX420978 |
| *Astrochapsa astroidea* | Luecking 24011 | Thailand | – | JX421445 | JX465278 |
| *Chroodiscus coccineus* | Herb. R. Luecking 2000 | Costa Rica | – | AF465441 | – |

Multi-locus (ITS, mtSSU and nuLSU) phylogenetic analysis was performed. The combined analysis included 70 sequences (Table 1) representing 18 in-group taxa and one out-group taxon. As many species as possible of *Chapsa* s. lat. were contained in our data matrix including the taxa that were similar in morphology or sequence to the new species and the two records. We blasted sequences of the three species in GenBank and selected sequence-similar taxa on a pre-determined cut-off.

The alignment was undertaken by applying MAFFT 7 with the option of L-INS-I (Katoh and Standley 2013). The three single-locus alignments were concatenated in PhyloSuite v1.2.2 (Zhang et al. 2020). The concatenated data matrix comprised 3188 nucleotide sites (nuLSU 1405 bp, ITS 647 bp and mtSSU 1136 bp). In order to check the consistency between the three loci, incongruence length difference test (ILD Test) was carried out using PAUP. The P value of ILD Test was 0.65 (>0.5), so the three loci were

suitable for polygenic phylogeny. Construction of the ML (Maximum Likelihood) tree was undertaken by applying RAxML v.8.2.12 (Stamatakis 2014) and implementing a GTRGAMMA model. For BI (Bayesian Inference) analysis, PartitionFinder 2 (Lanfear et al. 2017) was used to determine the best-fit model for each partition. For the nuLSU region, we used GTR+I+G, for ITS, GTR+G, and for mtSSU, HKY+I+G. BI analysis was performed with MrBayes 3.2.7 (Ronquist et al 2012). Markov Chain Monte Carlo (MCMC) chains were run for 200,000 generations, sampling every 100$^{th}$ generation, at which point, the average standard deviation of split frequencies was 0.001738. ML bootstrap values (BS) ≥ 75% and Bayesian posterior probabilities (PP) ≥ 0.95 were considered as significantly supported.

## Results and discussion

The BI and ML trees showed similar topologies and thus, only the BI tree was provided (Fig. 1). The three species were all monophyletic with a high support value: *C. mu-*

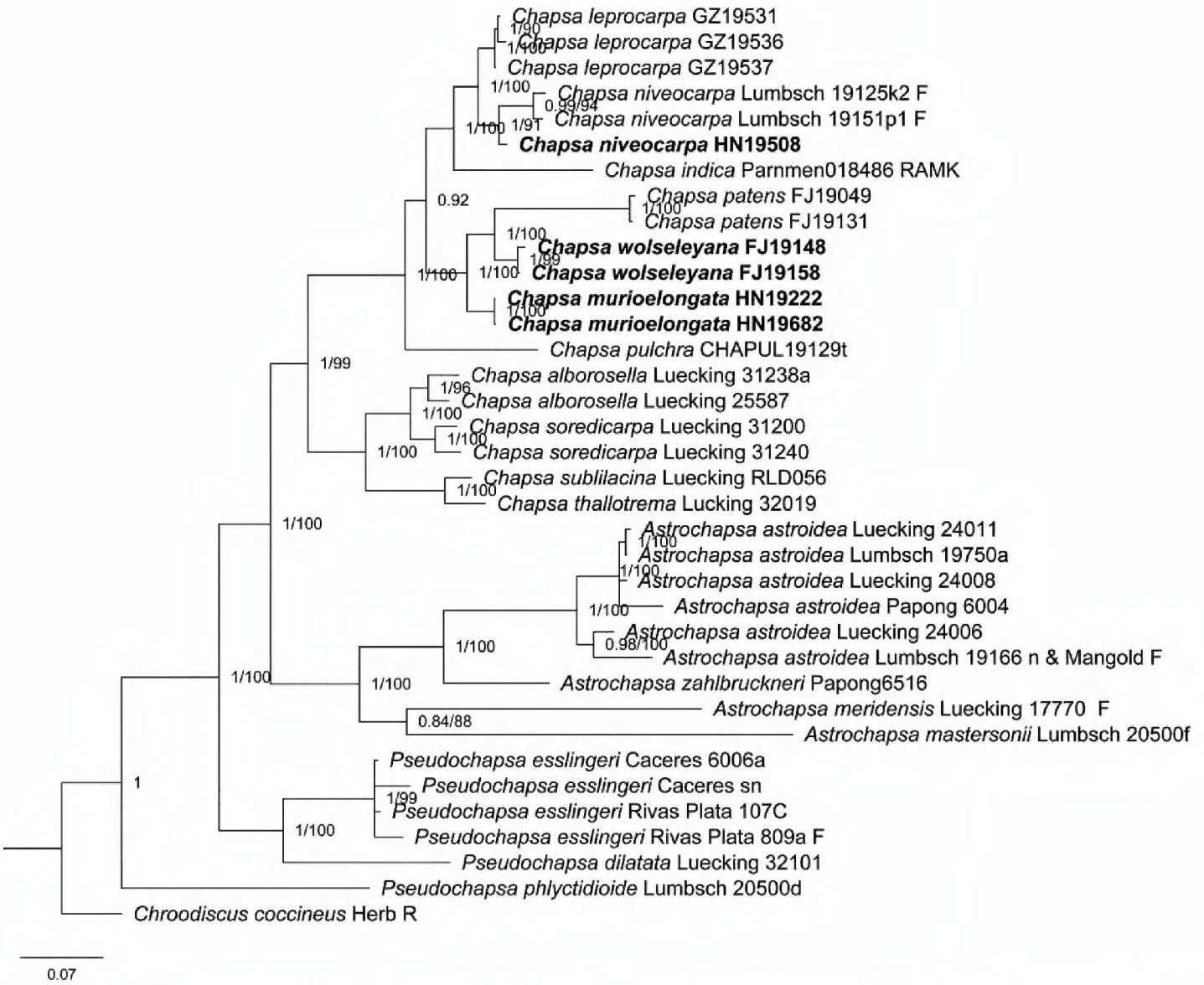


**Figure 1.** Bayesian phylogenetic tree generated from analysis of combined ITS, nuLSU and mtSSU. *Chroodiscus coccineus* is the out-group taxon. ML-bootstrap values/Bayesian posterior probabilities above 50% are written next to nodes.

*rioelongata* (100%, 1.00), *C. wolseleyana* (99%, 1.00) and *C. niveocarpa* (91%, 1.00). *Chapsa murioelongata* is sister to the clade consisting of *C. wolseleyana* and *C. patens* (Nyl.) Frisch. *Chapsa niveocarpa* HN19508 and *C. niveocarpa* Lumbsch form a well-supported clade and are sisters to *C. leprocarpa*.

## Taxonomy

### New species

***Chapsa murioelongata* M.Z. Dou & M. Li, sp. nov.**
Fungal Names: FN 570754
Figure 2

**Etymology.** The specific epithet *murioelongata* refers to the elongate, muriform ascospores.

**Type.** China. Hainan Province: Ledong County, Jianfengling National Forest Park, 18°42'39"N, 108°52'37"E, alt. 760 m, on bark, 09 Dec 2019, Y. H. Ju HN19222 (LCUF: holotype: HN19222; GenBank MW009102 for ITS and MW010271 for LSU).

**Description.** Thallus corticolous, crustose, olive-grey, surface dull, smooth to uneven, ecorticate. Apothecia erumpent, dispersed or two to four aggregated, rounded, 1–3 mm diam.; thalline margin lobed with white felt-like inner surface, lobes

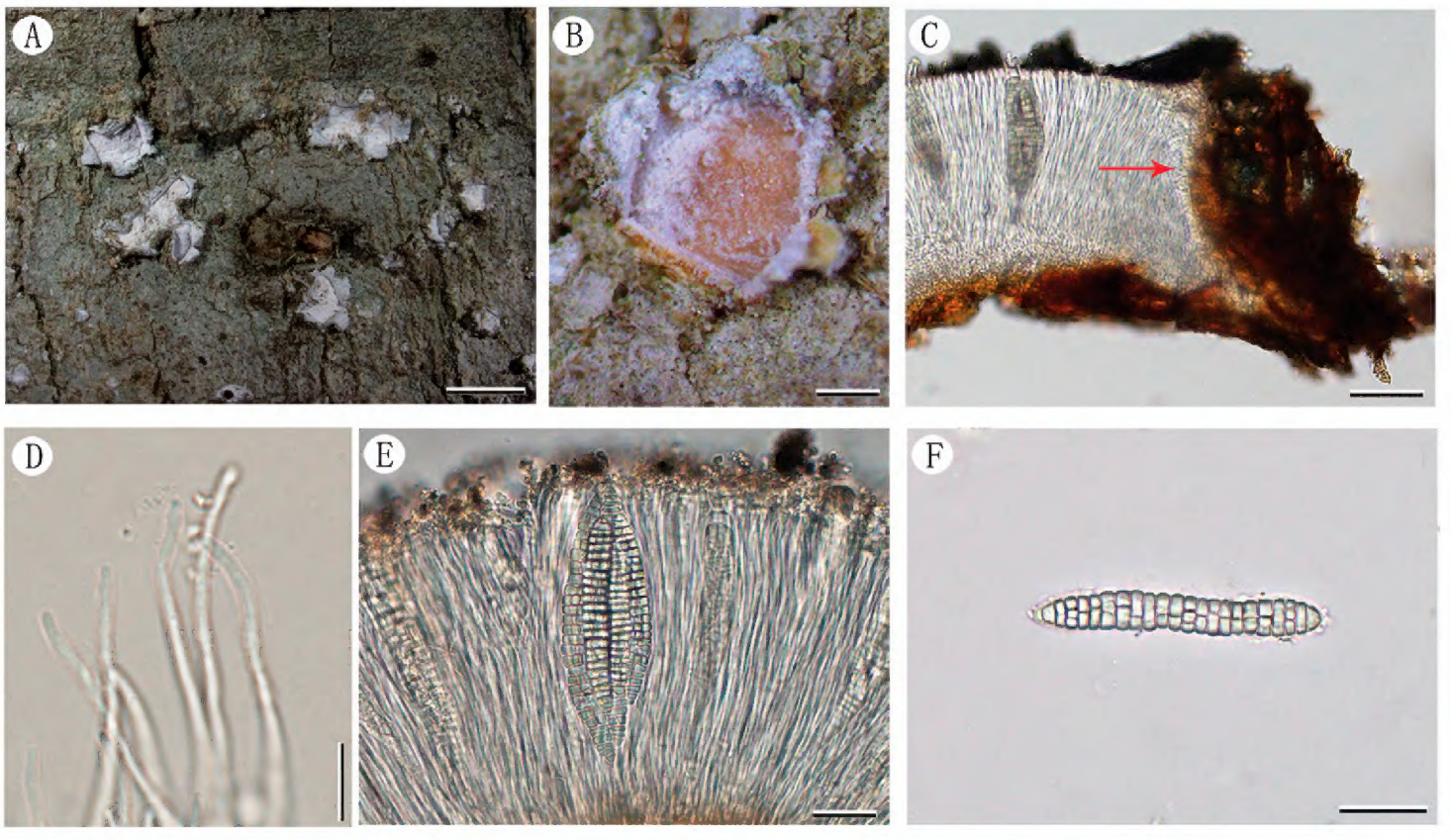


**Figure 2.** *Chapsa murioelongata* (LCUF HN19222) **A** habit of thallus with apothecia at different developmental stages **B** apothecium (the pruina of the disc partly scraped off) **C** section of apothecium with periphysoids (direction of arrow) **D** paraphyses **E** an ascus containing six ascospores **F** ascospore. Scale bars: 3 mm (**A**); 0.5 mm (**B**); 50 μm (**C**); 8 μm (**D**); 30 μm (**E**); 25 μm (**F**).

strongly backward curved; DISC flesh-coloured, covered by thick, white pruina. EXCIPLE 80–105 µm wide laterally, dark brown; EPIHYMENIUM 20–40 µm high, with coarse greyish granules; HYMENIUM clear, 110–170 µm high, non-amyloid; HYPOTHECIUM colourless, 10–30 µm high; PARAPHYSES simple, tips unbranched; PERIPHYSOIDES present, 5–30 µm long. ASCI 4–6 (8)-spored, clavate, 100–120 × 35–50 µm; ASCOSPORES hyaline, bacillary with rounded to subacute ends, submuriform with 20–25 transverse septa and 0–2 longitudinal septa per segment, 75–105 × 9.5–16 µm, non-halonate, I-. PYCNIDIA not observed.

**Chemistry.** Thallus K-, C-, PD-; no compounds detectable by TLC.

**Ecology and distribution.** On the bark in semi-exposed forest of Hainan Province.

**Additional specimens examined.** CHINA. Hainan Province: Changjiang County, Bawangling Nature Reserve, Yajia Scenic Area, 10°04'54"N, 109°07'04"E, alt. 810 m, on bark, 08 Dec 2019, Y. H. Ju HN19167 (LCUF); CHINA. Hainan Province: Lingshui County, Diaoluo Mountain, 18°43'35"N, 109°52'02"E, alt. 900 m, on bark, 14 Dec 2019, M. Li HN19682 (LCUF) (GenBank MW009103 for ITS and MW010269 for LSU).

**Note.** *Chapsa murioelongata* is characterised by its olive-grey thallus; white pruinose discs; distinct periphysoids; clear hymenium; 4–8-spored asci; submuriform ascospores with 20–25 transverse septa and 0–2 longitudinal septa per segment. *Chapsa microspora* Kalb, *C. asteliae* (Kantvilas & Vězda) Mangold, *Astrochapsa elongata* Poengs. & Lumbsch and *C. patens* are morphologically similar to the new species. *Chapsa microspora* can be distinguished from *C. murioelongata* by the smaller apothecia (0.6–1.2 mm diam.), transversely septate and smaller ascospores (7–9 × 4 µm) (Lumbsch et al. 2011). *Chapsa asteliae* differs in amyloid and shorter ascospores (30–80 µm) (Kantvilas and Vězda 2000; Mangold 2008). *Astrochapsa elongata* differs from *C. murioelongata* in having shorter ascospores (40–65 µm) and less longitudinal septa per segment (0–1) (Poengsungnoen et al. 2019). *Chapsa patens* differs from *C. murioelongata* chiefly in the single-spored asci and broader ascospores (22–35 µm) (Frisch et al. 2006).

Blast searches of nuLSU sequences indicate *Chapsa murioelongata* has close affinities with *C. patens* (98.36% identity), *C. wolseleyana* (95.63% identity), *C. leprocarpa* (91.97% identity) and *C. indica* (90.81% identity), so all these species were included in the phylogenetic analyses. *Chapsa murioelongata* was well separated from any other species in the tree and strongly supported as the monophyletic (PP = 1; ML = 100%).

## New records

### *Chapsa wolseleyana* Weerakoon, Lumbsch & Lücking, in Weerakoon, Rivas Plata, Lumbsch & Lücking, Lichenologist 44(3): 377 (2012)

Figure 3

*Astrochapsa wolseleyana* (Weerakoon, Lumbsch & Lücking) Parnmen, Lücking & Lumbsch, in Parnmen et al., PLoS ONE 7(12): 10 (2012)

**Description.** THALLUS crustose, corticolous, grey-brown, surface dull to slightly shiny, uneven, fissured. APOTHECIA erumpent, dispersed, sometimes two or three fused, most-

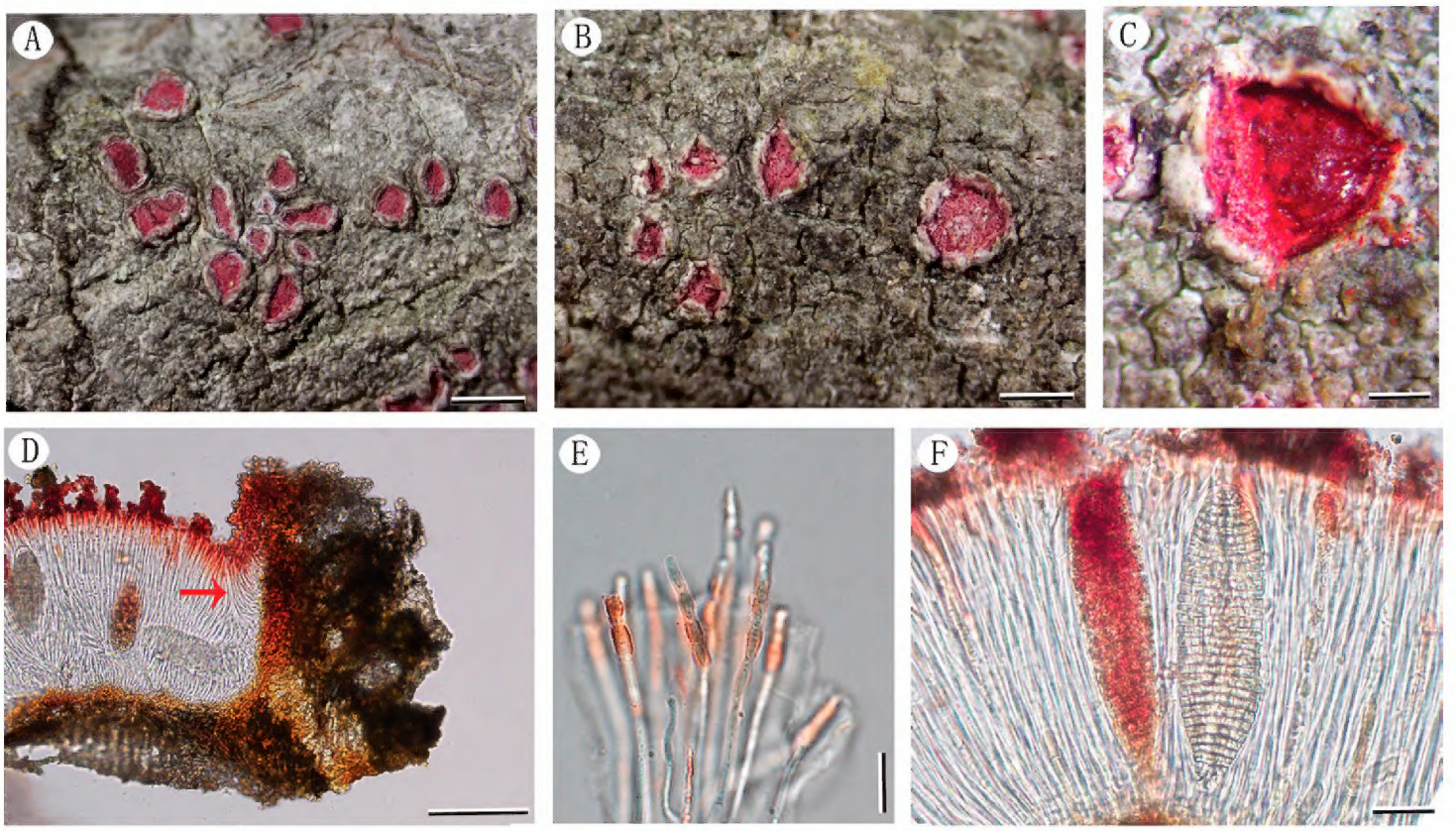


**Figure 3.** *Chapsa wolseleyana* (LCUF FJ19148-b) **A** habit of thallus with apothecia **B** apothecia at different developmental stages **C** apothecium (part of pruina scraped off) **D** section of apothecium with periphysoids (direction of arrow) **E** paraphyses **F** young and mature ascospores. Scale bars: 1.5 mm (**A**); 1 mm (**B**); 0.25 mm (**C**); 120 μm (**D**); 10 μm (**E**); 25 μm (**F**).

ly rounded to seldom slightly angular, 0.7–1.2 mm diam.; THALLINE MARGIN raised to lobulate, lobes erected to recurved, inner part brown, covered with rose-red or white pruina; DISC exposed, rose-red, covered with thick, rose-red pruina. EXCIPLE fused, cupular, laterally 180–250 μm wide, yellowish-brown to brown; EPIHYMENIUM rose-red with granules, 20–50 μm high, K+ green; HYMENIUM 140–230 μm high, clear, colourless, non-amyloid; HYPOTHECIUM indistinct; PARAPHYSES septate, tips rose-red and moniliform with oval or rectangular cells; PERIPHYSOIDES present, 50–100 μm long. ASCI clavate, 1-spored, 110–135 × 35–50 μm; ASCOSPORES densely muriform, oblong-ellipsoid, with hemispherical to roundish ends, 105–130 × 30–45 μm, first reddish, becoming hyaline to slightly olive-brown at maturity, I-. PYCNIDIA not observed.

**Chemistry.** No substances detected by TLC but apothecial disc with pigment producing K+ yellow-green efflux, suggesting presence of isohypocrelline.

**Ecology and distribution.** Growing on bark exposed to wind and high light intensity in montane forests. Worldwide distribution: Sri Lanka (Weerakoon et al. 2012) and newly reported for China.

**Selected specimens examined.** CHINA. Fujian Province: Quanzhou City, Jiuxian Mountain, Reflecting Pool, 25°42'57"N, 118°07'14"E, alt. 1540 m, on bark, 5 Jul 2019, F.Y. Liu FJ19148-b (LCUF) (GenBank MW009106 for ITS, MW010270 for LSU and MW010279 for SSU); CHINA. Fujian Province: Quanzhou City, Jiuxian Mountain, Natural Observation Path, 25°42'44"N, 118°07'17"E, alt. 1460 m, on bark, 25 Jul 2019, F.Y. Liu FJ19158 (LCUF) (GenBank MW009078 for ITS, MW010273 for LSU and MW010277 for SSU). CHINA. Fujian Province: Quanzhou City, Jiuxian Mountain,

Reflecting Pool, 25°42'57"N, 118°07'14"E, alt. 1540 m, on bark, 25 Jul 2019, F.Y. Liu FJ19127-2, same locality, FJ19128-2, FJ19141-2 (LCUF).

**Note.** *Chapsa wolseleyana* is characterised by its grey-brown, uneven thallus, apothecia with raised to lobed thalline margin, rose-red discs with similar coloured pruina, rose-red epihymenium and paraphyses tips, distinct periphysoids, 1-spored asci, muriform ascospores, red when young and hyaline to olive-brown when old. Only a few species of *Chapsa* have pigmented discs and among them *C. rubropulveracea* Hale ex Mangold, Lücking & Lumbsch is morphologically most similar to *C. wolseleyana*, but its thallus is farinose and its ascospores are 8 per ascus, smaller (15–20 ×5–6 μm) and transversely septate (Lumbsch et al. 2011).

*Chapsa wolseleyana* was transferred to *Astrochapsa*, based on a phenotype-based analysis (not molecular phylogeny) (Parnmen et al. 2012). However, our phylogenetic analysis shows that this species belongs in *Chapsa*, rather than *Astrochapsa*. *Chapsa wolseleyana* was associated phylogenetically with a strongly-supported clade (100/1) with *C. patens*, but with sufficient distance to be considered a distinct species. In addition, the latter differs from *C. wolseleyana* in having larger pale brown apothecia (up to 2 mm diam.) with white pruina, unpigmented epihymenium and unpigmented paraphyses adspersed with fine greyish to brownish granules, hyaline ascospores (Frisch et al. 2006; Joshi et al. 2012; Joshi et al. 2018).

***Chapsa niveocarpa*** **Mangold in Mangold, Elix & Lumbsch, Flora of Australia, 57:654 (2009)**

Figure 4

**Description.** Thallus corticolous, crustose, pale grayish-green surface dull and fluctuating along the bark. Apothecia erumpent, solitary to fused, angular rounded to slightly elongate, 0.5–1.8 × 0.5–1.2 mm; thalline margin split and recurved, inside-with thick white pruina; disc exposed, yellowish-brown, covered by white pruina. Exciple laterally 12–75 μm wide, dark brown; epihymenium 10–20 μm high; hymenium 120–200 μm high, grey-brown, inspersed by granules, non-amyloid; hypothecium indistinct; paraphyses unbranched; tips distinctly thickened; periphysoides present, but obscured by granular inclusions. Asci 1-spored, clavate, 120–140 × 27–36 μm; ascospores densely muriform, with thick halo at both ends, oblong, hyaline, 115–135 × 25–34 μm, I-. Pycnidia not observed.

**Chemistry.** Thallus K-, C-, PD-; no compounds detectable by TLC.

**Ecology and distribution.** Growing on tree bark in tropical rainforests in altitudes ranging from 500 to 1100 m. Australia, Queensland (Mangold 2008); newly reported for China.

**Selected specimens examined.** China. Hainan Province: Wuzhishan City, Wuzhishan Nature Reserve, 18°54'13"N, 109°41'04"E, alt. 870 m, on bark, 12 Dec 2019,

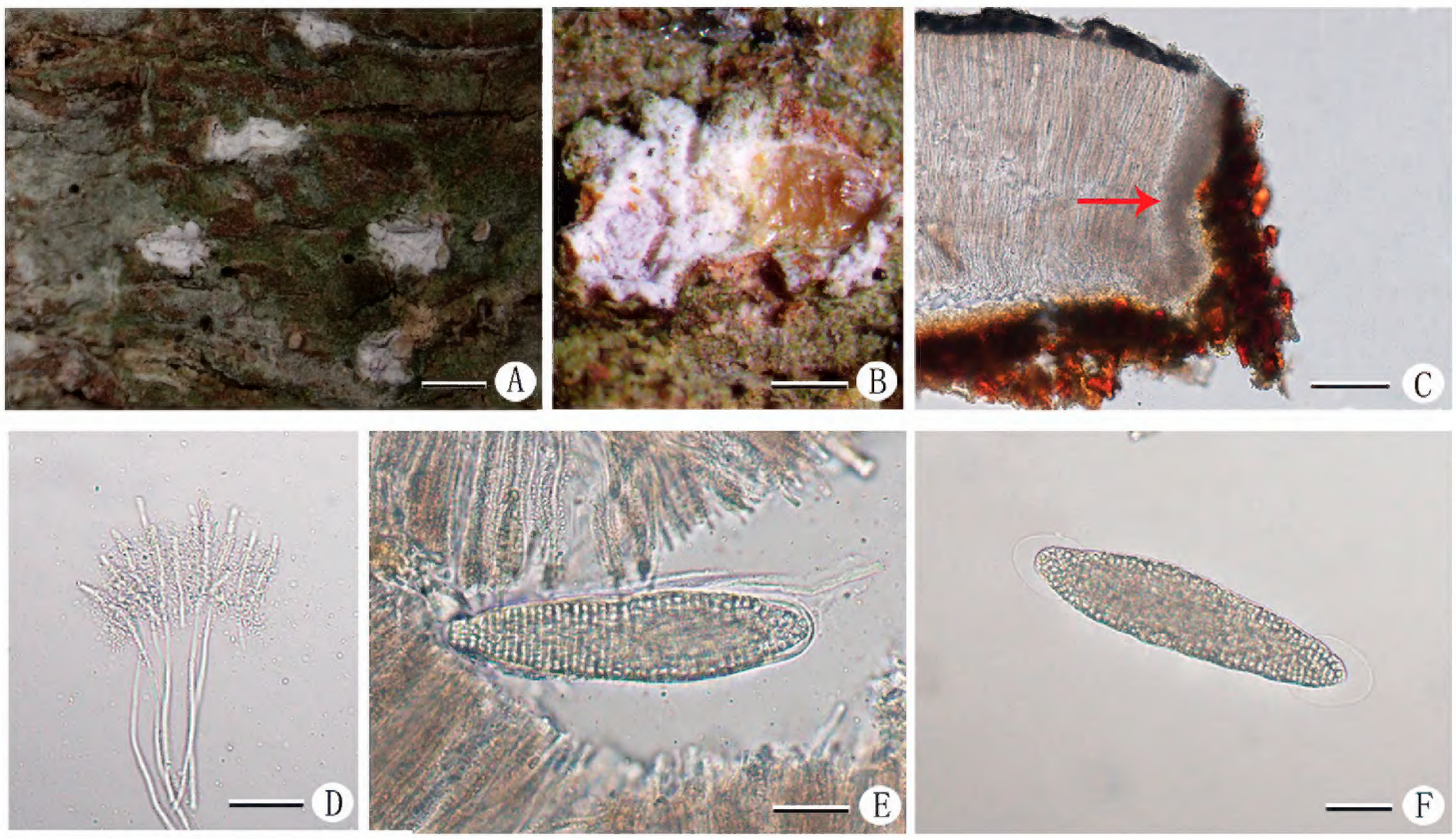


**Figure 4.** *Chapsa niveocarpa* (LCUF HN19508) **A** habit of thallus with apothecia **B** apothecium (part of pruina scraped off) **C** section of apothecium with periphysoids (direction of arrow) **D** paraphyses with hyaline granules **E** ascus **F** ascospore with halo. Scale bars: 1 mm (**A**); 0.5 mm (**B**); 50 μm (**C**); 25 μm (**D**); 30 μm (**E**); 25 μm (**F**).

M. Li HN19508 (LCUF) (GenBank MW009076 for ITS and MW010272 for LSU); China. Hainan Province: Wuzhishan City, Wuzhishan Nature Reserve, 18°53'13"N, 109°41'04"E, alt. 1020 m, on bark, 12 Dec 2019, M. Li HN19530 (LCUF); China. Hainan Province: Wuzhishan City, Wuzhishan Nature Reserve, 18°54'13'N, 109°41'04'E, alt. 870 m, on bark, 12 Dec 2019, M. Li HN19499 (LCUF); China. Hainan Province: Lingshui County, Diaoluo Mountain, 18°43'35"N, 109°52'02"E, alt. 900 m, on bark, 14 Dec 2019, M. Li HN19687 (LCUF); China. Hainan Province: Lingshui County, Diaoluo Mountain, 18°43'35"N, 109°52'02"E, alt. 900 m, on bark, 14 Dec 2019, M. Li HN19679 (LCUF).

**Note.** *Chapsa niveocarpa* is characterised by its crustose, pale greyish-green thallus; rounded to elongate apothecia, yellowish-brown discs with white pruina, obscured periphysoids, inspersed hymenium, 1-spored(rare 2-spored)ascus and muriform and hyalineascospores with halo. *Chapsa niveocarpa* is morphologically similar and phylogenetically related to *C. leprocarpa*, and both species occur on bark in tropical forests (Frisch 2006; Mangold 2008; Parnmen et al. 2012). *Chapsa leprocarpa* differs from *C. niveocarpa* in having a lower hymenium (100–130 μm) and smaller ascospores (up to 111 μm long) (Frisch 2006). The specimen (HN19508) we collected in China is allocated phylogenetically to a strongly-supported (1/91) clade with *C. niveocarpa*. The collections cited above are the first reports for China.

## Key to *Chapsa* in China

1 Disc with red pruina; ascospores 1/ascus, muriform, 105–135 × 30–50 μm. ..... ***C. wolseleyana***

– Disc with white pruina ..... **2**

2 Ascospores transversely septate; ascospores 4–8/ascus, 50–110 × 6–12 μm.... ..... ***C. indica***

– Ascospores (sub)muriform ..... **3**

3 Hamathecium inspersed; ascospores 1/ascus, 80–190 × 20–50 μm ..... ..... ***4***

– Hamathecium clear..... **5**

4 Ascospores 1/ascus, 80–190 × 20–50 μm..... ***C. niveocarpa***

– Ascospores 8/ascus, 40–50 × 11–15 μm..... ***C. mirabilis***

5 Asci 4–6 (8)-spored; acsospores oblong to cylindrical with rounded to subacute ends, submuriform with 20–25 transverse septa and 0–2 longitudinal septa per segment, 75–105 × 9.5–16 μm ..... ***C. murioelongata***

– Asci 4-spored; acsospores oblong to slightly ellipsoid, with roundish ends, 60–130 × 20–40 μm..... ***C. leprocarpa***

## Acknowledgements

This study was supported by the National Natural Science Foundation of China (31750001), Doctoral Initiation Fund of Liaocheng University (318051813) and Research Fund of Liaocheng University (318012011).

## References

Dou MZ, Wu XH, Li M, Zhao X, Jia ZF (2018) *Gyalecta caudiospora* sp. nov. from China. Mycotaxon 133(4): 721–727. https://doi.org/10.5248/133.721

Frisch A (2006) Contribution towards a new systematic of the lichen family Thelotrematáceae. I. The lichen family Thelotrematáceae in Africa. Bibliotheca Lichenologica 92: 1–370.

Frisch A, Kalb K (2009) *Chapsa* species (Thelotrematáceae) from Brazil. Bibliotheca Lichenologica 99: 133–142.

Gardes M, Bruns TD (1993) ITS primers with enhanced specificity for Basidiomycetes – application to the identification of mycorrhiza and rusts. Molecular Ecology 2: 113–118. https://doi.org/10.1111/j.1365-294X.1993.tb00005.x

Jia ZF, Lücking R (2017) Resolving the genus *Phaeographina* Müll. Arg. in China. Mycokeys 21(1): 13 –32.

Jia ZF, Wei JC (2016) Flora Lichenum Sinicorum, vol. 13. – Ostropales (I) – Graphidaceae 1. Science Press, Beijing, 21–22.

Joshi S, Upreti DK, Nayaka S (2012) The lichen genus *Chapsa* (Graphidaceae) in India. Mycotaxon 120: 23–24, 27, 31, 32. https://doi.org/10.5248/120.23

Joshi S, Upreti DK, Divakar PK, Lumbsch HT, Lücking R (2018) A re-evaluation of thelotremoid Graphidaceae (lichenized Ascomycota: Ostropales) in India. The Lichenologist 50(6): 634–639. https://doi.org/10.1017/S0024282918000439

Kalb J, Kalb K (2017) New lichen species from Thailand, new combinations and new additions to the Thai lichen biota. Phytotaxa 332 (2): 141–156. https://doi.org/10.11646/phytotaxa.332.2.2

Kantvilas G, Vězda A (2000) Studies on the lichen family Thelotremataceae in Tasmania. The genus *Chroodiscus* and its relatives. Lichenologist 32(4): 325–357. https://doi.org/10.1006/lich.2000.0274

Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Molecular Biology and Evolution 30: 772–780. https://doi.org/10.1093/molbev/mst010

Kraichak E, Parnmen S, Lücking R, Lumbsch HT (2013) *Gintarasia* and *Xalocoa*, two new genera to accommodate temperate to subtropical species in the predominantly tropical Graphidaceae (Ostropales, Ascomycota). Australian Systematic Botany 26: 466–474. https://doi.org/10.1071/SB13038

Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B (2017) PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. Molecular Biology and Evolution 34(3): 772–773. https://doi.org/10.1093/molbev/msw260

Lima EL, Maia LC, Martins MCB, da Silva NL, Lücking R, da Silva Cáceres ME (2019) Five new species of Graphidaceae from the Brazilian Northeast, with notes on *Diorygma alagoense*. The Bryologist 122(3): 414–422. https://doi.org/10.1639/0007-2745-122.3.414

Lumbsch HT, Ahti T, Altermann S, De Paz GA, Aptroot A, Arup U et al. (2011) One hundred new species of lichenized fungi: a signature of undiscovered global diversity. Phytotaxa 18: 36–40. https://doi.org/10.11646/phytotaxa.18.1.1

Mangold A (2008) Taxonomic studies on members of Thelotrematoid Ostropales (Lichenized Ascomycota) in Australia. PhD Thesis, University of Duisburg, Essen.

Massalongo AB (1860) Esame comparativo di alcune genere di licheni. Atti dell´Istituto Veneto Scienze 5: 247–276.

Orange A, James PW, White FJ (2010) Microchemical methods for the identification of lichens. British Lichen Society, London, 101 pp.

Papong K, Boonpragob K, Mangold A, Divakar PK, Lumbsch HT (2010) Thelotremoid lichen species recently described from Thailand: a re-evaluation. The Lichenologist 42(2): 131–137. https://doi.org/10.1017/S0024282909990405

Parnmen S, Lücking R, Lumbsch HT (2012) Phylogenetic classification at generic level in the absence of distinct phylogenetic patterns of phenotypical variation: a case study in Graphidaceae (Ascomycota). PLoS ONE 7(12): e51392. https://doi.org/10.1371/journal.pone.0051392

Parnmen S, Cáceres MES, Lücking R, Lumbsch HT (2013) *Myriochapsa* and *Nitidochapsa*, two new genera in Graphidaceae (Ascomycota: Ostropales) for chroodiscoid species in the *Ocellularia* clade. The Bryologist 116(2): 127–133. https://doi.org/10.1639/0007-2745-116.2.127

Poengsungnoen V, Buaruang K, Vongshewarat K, Sangvichien K, Boonpragob K, Mongkolsuk P, Lumbsch HT (2019) Three new crustose lichens from Thailand. The Bryologist 122(3): 451–456. https://doi.org/10.1639/0007-2745-122.3.451

Rivas Plata R, Lücking R, Lumbsch HT (2011) A new classification for the family Graphidaceae (Ascomycota: Lecanoromycetes: Ostropales). Fungal Diversity 52(1): 107–121. https://doi.org/10.1007/s13225-011-0135-8

Rivas Plata R, Lücking R, Sipman HJM, Mangold A, Klab K, Lumbsch HT (2010) A worldwide key to the thelotremoid Graphidaceae, excluding the Ocellularia-Myriotrema-Stegobolus clade. The Lichenologist 42(2): e139185.

Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Hohna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61: 539–542. https://doi.org/10.1093/sysbio/sys029

Sipman HJM, Lücking R, Aptroot A, et al. (2012) A first assessment of the Ticolichen biodiversity inventory in Costa Rica and adjacent areas: the thelotremoid Graphidaceae (Ascomycota: Ostropales). Phytotaxa 55: 1–214. https://doi.org/10.11646/phytotaxa.55.1.1

Stamatakis A (2014) RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30(9): 1312–1313. https://doi.org/10.1093/bioinformatics/btu033

Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplifed ribosomal DNA from several *cryptococcus* species. Journal of Bacteriology 172(8): 4238–4246. https://doi.org/10.1128/JB.172.8.4238-4246.1990

Weerakoon G, Rivas Plata, Lumbsch HT, Lücking R (2012) Three new species of *Chapsa* (lichenized Ascomycota: Ostropales: Graphidaceae) from tropical Asia. The Lichenologist 44(3): 373–379. https://doi.org/10.1017/S0024282911000892

Wei JC (2020) The enumeration of lichenized fungi in China. China Forestry Publishing House, Beijing, 606 pp.

White T, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Snisky JJ, White TJ (Eds) PCR Protocols: a guide to methods and applications. Academic Press, New York, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1

Wijayawardene NN, Hyde KD, Rajeshkumar KC, Hawksworth DL, Madrid H, Kirk PM, Braun U, Singh RV, Crous PW, Kukwa M, Lücking R, Kurtzman CP, Yurkov A, Haelewaters D, Aptroot A, Lumbsch HT, Timdal E, Ertz D, Etayo J, Phillips AJL, Groenewald JZ, Papizadeh M, Selbmann L, Dayarathne MC, Weerakoon G, Jones EBG, Suetrong S, Tian Q, Castañeda-Ruiz RF, Bahkali AH, Pang KL, Tanaka K, Dai DQ, Sakayaroj J, Hujslová M, Lombard L, Shenoy BD, Suija A, Maharachchikumbura SSN, Thambugala KM, Wanasinghe DN, Sharma BO, Gaikwad S, Pandit P, Zucconi L, Onofri S, Egidi E, Raja HA, Kodsueb R, Cáceres MES, Pérez-Ortega S, Fiuza PO, Monteiro JS, Vasilyeva LN, Shivas RG, Prieto M, Wedin M, Olariaga I, Lateef AA, Agrawal Y, Fazeli SAS, Amoozegar MA, Zhao GZ, Pfliegler WP, Sharma G, Oset M, Abdel-Wahab MA, Takamatsu S, Bensch K, de Silva NI, Kesel AD, Karunarathna A, Boonmee S, Pfister DH, Lu YZ, Luo ZL, Boonyuen N, Daranagama DA, Senanayake IC, Jayasiri SC, Samarakoon MC, Zeng XY, Doilom M, Quijada L, Rampadarath S, Heredia G, Dissanayake AJ, Jayawardana RS, Perera RH, Tang LZ, Phukhamsakda C, Hernández-Restrepo M, Ma X, Tibpromma S, Gusmao LFP,

Weerahewa D, Karunarathna SC (2017) Notes for genera: Ascomycota. Fungal Diversity 86: e96. https://doi.org/10.1007/s13225-017-0386-0

Xu LL, Wu QH, Wang QD, Jia ZF (2016) *Chapsa* (Graphidaceae, Ostropales), A lichen genus new to China. Journal of Tropical and Subtropical Botany 24(5): 495–498.

Zhang D, Gao FL, Jakovli I, Zou H, Wang GT (2020). Phylosuite: an integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. Molecular Ecology Resources 20(1): 348–355. https://doi.org/10.1111/1755-0998.13096

Zoller S, Scheidegger C, Sperisen C (1999) PCR primers for the amplification of mitochondrial small subunit ribosomal DNA of lichen-forming ascomycetes. Lichenologist 31: 511–516. https://doi.org/10.1006/lich.1999.0220